

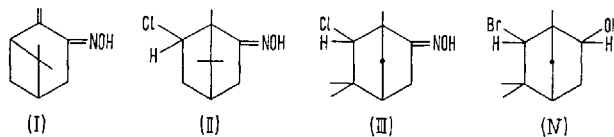
noch nicht beschriebenen 6-Chlor-Campheroxims (II) entspricht. Über chemischen Abbau und Stereochemie dieser Verbindung wird gesondert berichtet.

6-Chlor-Campheroxim entsteht auch als Nebenprodukt beim Einwirken von Nitrosylchlorid auf α -Pinen sowie bei der Reaktion von Chlorwasserstoff mit Pinocarvonoxim. Die hierbei erzielten Ausbeuten sind jedoch nur gering.

Im Zuge der Konstitutionsaufklärung des 6-Chlor-Campheroxims stellten wir uns zu Vergleichszwecken «Pinocarvonoximhydrochlorid»¹ her.

Das NMR-Spektrum dieser Verbindung zeigt neben den Signalen für die Methylgruppen (1,0, 1,16 und 1,26 ppm) und dem Singulett des Oxim-Protons bei 8,7 ppm ein weiteres Singulett bei 3,7 ppm. Das Auftreten dieses Signals sowie das Fehlen einer charakteristischen Bande für die Partialstruktur $\text{Cl}-\overset{\text{H}}{\underset{|}{\text{C}}}-\text{CH}_3$ ² schliesst die von TREIBS¹

vorgeschlagene Struktur eines Pinan-Derivates aus. Eine Camphan-Struktur kommt ebenfalls nicht in Betracht. Dagegen lässt sich das Singulett gut durch das Isofenchan-Grundgerüst erklären. Es entspricht danach dem Proton



C₆, das durch den stark negativen Cl-Substituenten am gleichen C-Atom nach tieferem Feld verschoben wird. Der Verbindung von TREIBS¹ muss daher die Struktur des 6-Chlor-Isofenchonoxims (III) zugeschrieben werden. Unsere Ergebnisse ergänzen Feststellungen von HARTSHORN und WALLIS³, wonach beim Einwirken von Bromwasserstoff auf Pinocarveol ebenfalls ein Derivat des Isofenchans, und zwar der 6-Brom-Isofenchylalkohol (IV) entsteht.

Summary. The nitrosochloride of pinene dissolved in benzene will be transformed quantitatively by adsorbing to SiO₂ to the not yet described 6-chlor-campheroxim. This compound is also a by-product in the reaction of α -pinene with nitrosylchloride.

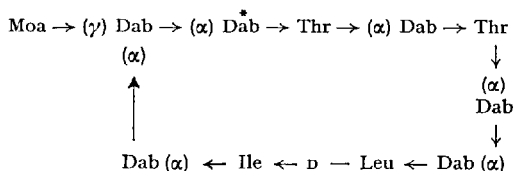
C. H. BRIESKORN und W. RUSSOW⁴

Institut für Pharmazie und Lebensmittelchemie der Universität, 87 Würzburg (Deutschland), 21. Februar 1968.

- 1 W. TREIBS, M. MÜHLSTÄDT, R. MEGGES und J. KLOTZ-HERDMANN, Justus Liebigs Annln Chem. 634, 118 (1960).
- 2 L. M. JACKMAN, *Application of NMR in Organic Chemistry* (London-Oxford-New York-Paris 1959, Pergamon Press 1959).
- 3 M. P. HARTSHORN and A. F. A. WALLIS, *Chem. Ind.* 1878 (1963).
- 4 Teil der Dissertation W. Russow, Univ. Würzburg 1967.

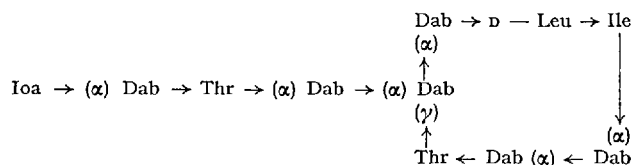
Chemical Structure of Circulin B

In 1949 circulins A and B were isolated from the culture fluid of *Bacillus circulans* Q-19¹. In 1958 KOFFLER and KOBAYASHI² reported that the chemical structures of these antibiotics are cyclic decapeptides, and circulin B has a similar structure to circulin A, with the exception that the Moa³ is attached through the γ -amino group of the Dab marked by an asterisk in the structure of circulin A (Scheme 1).



Scheme 1. Chemical structure for circulin B by KOFFLER and KOBAYASHI²

This seemed to be very unusual in the view of our results obtained in studies of the chemical structures of polymyxin series antibiotics⁴. Namely, colistins A and B, polymyxins B₁ and B₂, or polymyxins D₁ and D₂ differ from each other only in the nature of their fatty acid components. The result of our re-examination now proves the structure of circulin B to be that shown in Scheme 2.



Scheme 2. Full structure for circulin B

Circulin B was isolated from the culture fluid of *B. circulans* ATCC 14040 by ion exchange chromatography (Amberlite IRC-50, H⁺-form) and by countercurrent distribution. The amino acid composition of the purified circulin B was confirmed as L-Thr:D-Leu:L-Ile:L-Dab in the molar ratio of 2:1:1:6. The methyl ester derivative of the fatty acid isolated from the acid hydrolysate of circulin B was found to be isooctanoic acid methyl ester by gas chromatography. This result conflicted with the report of KOFFLER and KOBAYASHI² that circulin B

- 1 F. J. MURREY, P. A. TETRAUL, O. W. KAULMANN, H. KOFFLER, D. H. PETERSON and D. R. COLINGSWORTH, *J. Bact.* 57, 305 (1949). - D. H. PETERSON and L. N. REINEKE, *J. biol. Chem.* 181, 95 (1949). - J. H. DOWLING, H. KOFFLER, H. C. REITZ, D. H. PETERSON and P. A. TETRAUL, *Science* 116, 147 (1952).
- 2 H. KOFFLER and T. KOBAYASHI, *Abstr. Div. Biol. Chem. Ann. Chem. Soc., San Francisco*, April 13-18 (1958).
- 3 The following abbreviations are used throughout this paper; Moa, (+)-6-methyloctanoyl residue; Ioa, isooctanoyl residue; Dab, α,γ -diaminobutyric acid residue; DNP, 2,4-dinitrophenyl residue; $\rightarrow$ =C to N bond in -CO-NH; $\rightarrow$ (alpha) Dab = $\rightarrow$ Dab; $\rightarrow$ (gamma) Dab = $\rightarrow$ Dab.
- 4 T. SUZUKI, H. INOUE, K. FUJIKAWA and Y. SUKETA, *J. Biochem.* 54, 25 (1963). - T. SUZUKI, H. INOUE, K. FUJIKAWA and S. NAGASAWA, *J. Biochem.* 54, 173 (1963). - T. SUZUKI, K. HAYASHI and K. FUJIKAWA, *J. Biochem.* 54, 412 (1963). - T. SUZUKI and K. FUJIKAWA, *J. Biochem.* 56, 182 (1964). - T. SUZUKI, K. HAYASHI, K. FUJIKAWA and K. TSUKAMOTO, *J. Biochem.* 56, 335 (1964). - K. FUJIKAWA, Y. SUKETA, K. HAYASHI and T. SUZUKI, *Experientia* 21, 307 (1965). - K. HAYASHI, Y. SUKETA, K. TSUKAMOTO and T. SUZUKI, *Experientia* 22, 354 (1966).

contains 6-methyloctanoic acid. By applying polymyxin acylase⁶, which was isolated from soil bacteria, to circulin B, deacyl circulin B was obtained. From the acid hydrolysate of DNP-derivative of deacyl circulin B, di-DNP-Dab was detected as the *N*-terminal amino acid, and the molar ratios of free amino acids in the acid hydrolysate of DNP-derivative of circulin B were found to be Thr:Leu:Ile:Dab = 2:1:1:1. These results suggested that circulin B does not have the structure proposed by KOFFLER and KOBAYASHI².

Then, using our technique with Nagarse³ (subtilo peptidase A, EC 3.4.4.16), we obtained the following fragments: (I) Ioa $\rightarrow$ (α)Dab $\rightarrow$ Thr, (II) Ioa $\rightarrow$ (α)Dab $\rightarrow$ Thr $\rightarrow$ (α)Dab, (III) Dab, (IV) a cyclic heptapeptide having the molar ratios of amino acids of Thr:Leu:Ile:Dab = 1:1:1:4. Applying the SANGER'S DNP method to IV we detected α -DNP-Dab, which could not be found in the completely dinitrophenylated circulin B. The AKABORI'S hydrazinolysis method⁶ failed to find the carboxy terminal amino acid of peptide IV. The Rf value of IV was identical with that of the cyclic heptapeptide obtained from circulin A by the enzymatic hydrolysis using Nagarse, by comparison with their Rf values in paper chromatography. In addition, considering the chemical

structures of other polymyxin series antibiotics⁴, the amino acid sequence of peptide IV was found to be cyclo(γ)Dab $\rightarrow$ (α)Dab $\rightarrow$ Leu $\rightarrow$ Ile $\rightarrow$ (α)Dab $\rightarrow$ (α)Dab $\rightarrow$ Thr $\rightarrow$.

On the basis of these results, circulin B has a similar structure to circulin A as shown in Scheme 2 and only differs from circulin A in having Ioa in place of Moa.

Zusammenfassung. Es wird über die Strukturaufklärung von Circulin B, einem Antibiotikum aus *Bacillus circulans* ATCC 14040, berichtet, dem die Formel des Schemas 2 zukommt.

K. HAYASHI, Y. SUKETA
and T. SUZUKI

*Institute for Protein Research, Osaka
University, Osaka (Japan), 26 March 1968.*

⁵ T. SUZUKI, Y. KIMURA and K. IWADARE, Jap. Patent Gazette 12, 101 (1966).

⁶ S. AKABORI, K. OHNO and K. NARITA, Bull. Chem. Soc. Japan 25, 214 (1952).

The Synthesis of a Natural Dinucleoside Phosphate Derivative with the Aid of a Purine Cyclonucleoside

In a previous communication¹ we described the use of an 8,5'-*o*-purine cyclonucleoside for the synthesis of a dinucleoside phosphate. As this procedure yielded a product having a hydroxyl group at position 8 of the purine moiety, it was of interest to study the opening of the anhydro linkage of an 8,5'-*S*-purine cyclonucleoside by the attack of a nucleoside phosphate anion. Such an approach would lead to the formation of a dinucleoside phosphate with a mercapto group at position 8 of the purine nucleus, which could be removed by treatment with Raney Ni, thus resulting in a product having no unnatural substituents.

8,5'-Anhydro-8-mercapto-2'-*o*-mesylguanosine² (200 mg) was, therefore, refluxed with 1.2 equivalents of tri-*n*-butyl ammonium 3'-uridylylate in dry DMF for 14 h. The dinucleoside phosphate, uridyl-(3'-5')-8-mercapto-2'-*o*-mesylguanosine (I), was isolated by preparative paper chromatography on Whatman 3 mm paper³ in 41% yield. Rf, 0.37; $\lambda_{\max}$, 0.1 N HCl-265.5, 295 (sh) nm; 0.1 N NaOH 268, 285 (sh) nm. Formic acid hydrolysis of this product gave uracil and 8-mercaptoguanine. Paper chromatographic analysis of the snake venom phosphodiesterase

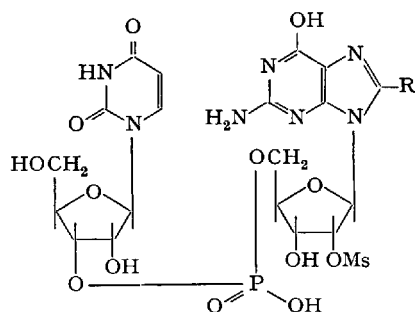
hydrolysate showed that uridine and another substance, presumably 8-mercapto-2'-*o*-mesylguanosine-5'-phosphate, were formed. That the methyl sulphonate group at the 2' position in the purine cyclonucleoside was not displaced (or displaced only to a very small extent) by the phosphate anion was shown by the presence of absorption at 1170 cm⁻¹ (Nujol) in the IR-spectrum of I. This is consistent with an observation made earlier that it is difficult to remove this group².

Uridyl-(3'-5')-8-mercapto-2'-*o*-mesylguanosine (I) was refluxed with Raney Ni in aqueous ethanol (1:1) for 3.5 h. Filtration of the catalyst and evaporation of the solvent yielded uridyl-(3'-5')-2'-*o*-mesylguanosine (II) in 75% yield, Rf 0.40³; $\lambda_{\max}$, 0.1 N HCl-260, 0.1 N NaOH-262 nm. This product contained sulphur, had mesyl absorption in the IR and yielded guanine and uracil on formic acid hydrolysis⁴.

Zusammenfassung. Die Behandlung von 8-5'-Anhydro-8-mercapto-2'-*o*-mesylguanosin mit (1,2 Äquivalent) Tri-*n*-butylammonium-uridylat in siedendem DMF lieferte das Dinucleosidphosphat Uridyl-(3'-5')-8-mercapto-2'-*o*-mesylguanosin. Weitere Behandlung mit Raney Ni führte zum Uridyl-(3'-5')-2'-*o*-mesylguanosin.

P. C. SRIVASTAVA, K. L. NAGPAL
and M. M. DHAR

*Central Drug Research Institute, Lucknow (India),
4 March 1968.*



I. R = SH; II, R = H

¹ K. L. NAGPAL and M. M. DHAR, Tetrahedron Lett. 47 (1968).

² M. IKEHARA, H. TADA and K. MUNAYAMA, Chem. pharm. Bull., Tokyo 13, 639 (1965).

³ Descending paper chromatography, solvent: *n*-butanol-acetic acid-water (4:1:5).

⁴ Communication No. 1252 from the Central Drug Research Institute.